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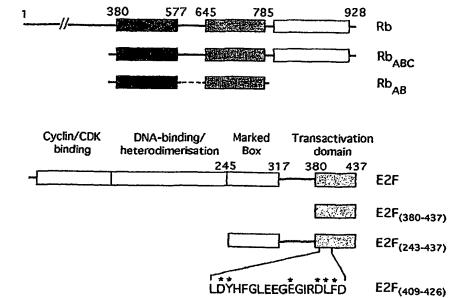
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(54) Title: STRUCTURE OF A COMPLEX OF RETINOBLASTOMA PROTEIN BOUND TO E2F, AND USES THEREOF



(57) Abstract: The present invention provides the crystal structure of  $pRb/E2F_{(409-426)}$  as well as uses of the structure in identifying agents which modulate the binding between pRb and E2F and/or a  $pRb/E2F_{(409-426)}$  complex, and thus are useful as pharmaceutical agents in the prevention or treatment of proliferative diseases.

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STRUCTURE OF A COMPLEX OF RETINOBLASTOMA PROTEIN BOUND TO E2F, AND USES THEREOF

The present invention relates to the crystal structure of pRb/E2F<sub>(409-426)</sub> as well as uses of the structure in identifying agents which modulate the binding between pRb and E2F and/or a pRb/E2F<sub>(409-426)</sub> complex, and thus are useful as pharmaceutical agents in the prevention or treatment of proliferative diseases.

The retinoblastoma tumour suppressor protein (pRb) regulates the cell cycle, sponsors differentiation and restrains apoptosis. Dysfunctional pRb is thought to be necessary for the development of most human malignancies.

pRb controls the cell cycle and apoptosis by acting as a negative regulator of transcription. It is now established that the growth-inhibitory effects of pRb are dependent on its regulation of the E2F family of transcription factors whose activity is necessary for the expression of genes involved in the G1 to S transition of the cell cycle and DNA replication. The transcriptional repression exerted by pRb over E2F responsive promoters involves at least three, distinct mechanisms. By binding to the transcriptional activation domain of E2F, pRb prevents it from recruiting components of the transcriptional apparatus and, once tethered to E2F promoters, pRb interacts with, and represses, other nearby transcription factors. Finally, pRb recruits protein factors to E2F promoters, such as histone deacetylases (HDACs) and histone methyltransferases (HMTases) that negatively regulate transcription by altering chromatin structure.

In addition to regulating entry into S-phase, it is thought that pRb is important in protecting differentiating cells from apoptosis. Certainly in many types of tissue, loss of pRb leads to apoptosis. This and other data has led to a model whereby the anti-apoptotic activity of pRb is mediated by its repression of certain E2F-dependent promoters. Unrepressed E2F is able to drive apoptosis by both p53-dependent and p53-independent mechanisms.

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Although inactivation of the pRb pathway is thought to be widely involved in cellular transformation, there are examples of tumours where over-expression of functional pRb appears to be detrimental to successful clinical treatment. For example, adenocarcinoma of the pancreas is the fifth most common cause of cancer-related death in the Western world. It is particularly resistant to currently available forms of chemotherapy and radiation therapy. It is thought that this malignancy is able to evade apoptosis induced by treatment with chemotherapeutic drugs because of over-expression of pRb. It seems plausible that the protective effect of pRb over-expression against apoptosis is mediated by E2F. By blocking transcriptional activation by E2F, over-expression of pRb appears to render pancreatic cancer cells insensitive to chemotherapy.

As many of the anti-tumourigenic properties of pRb are mediated by its regulation of the E2F transcription factors, it would be beneficial to have a crystal structure of the pRb-binding fragment of E2F (E2F<sub>(409-426)</sub>) in complex with the tumour suppressor protein. Such detailed knowledge of the molecular interactions between E2F and the A/B interface of pRb would enable the development of compounds that bind to pRb and inhibit complex formation. Such a compound, administered in parallel with conventional chemotherapy, would offer a means of enhancing treatment of proliferative diseases such as pancreatic cancer and perhaps related diseases.

Accordingly, the present invention provides the crystal structure of the primary pRb-binding fragment of E2F (E2F<sub>(409-426)</sub>) in complex with the tumour suppressor protein pRb. The structure shows how E2F<sub>(409-426)</sub> binds at the interface of the A and B domains of the pocket of pRb making extensive interactions with conserved residues from both.

In order to address the regulation of the E2F transcription factor by pRb, the present inventors have determined the crystal structure of the complex of pRb<sub>AB</sub> bound to the

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minimal binding region of E2F, namely E2F<sub>(409-426)</sub>. The structure has important implications for the understanding of pRb/E2F function. The studies have quantified the contribution of the principal interaction made by E2F through residues 409-426 with pRb as well as that of a secondary interaction involving the marked box region of E2F. In both cases these interactions are with the pocket region of the tumour suppressor protein pRb.

The analysis of the crystal structure of pRb/E2F<sub>(409-426)</sub> suggests that E2F<sub>(409-426)</sub> acts as a sensor of the structural integrity of the pRb pocket. Accordingly, cells in many tissues should be protected against deleterious mutations in pRb because they will sponsor increased E2F transcriptional activation, and thus apoptosis. It seems particularly intriguing, therefore, that all tumour derived pRb mutants fail to bind to E2F suggesting that an intense selectionary pressure operates in many types of tissue in favour of cells which harbour defects in apoptosis once they have lost normal pRb function. Perhaps the most notable exception to this process occurs in retinal cells, which are able to survive for some time with loss of pRb without acquiring other genetic alterations. Indeed, it has been suggested that these particular cells are distinguished by their ability to acquire survival signals from neighbouring cells and thus give rise to the eponymous retinoblastomas.

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According to a first aspect, the present invention provides a crystal structure of the pRb/E2F<sub>(409-426)</sub> complex, characterised by the atomic co-ordinates of Annex 1.

Preferably the interactions between E2F<sub>(409-426)</sub> and pRb comprise one or more of the following interactions:

E2F <sub>(409-426)</sub> residue	pRb residue
Leu <sub>409</sub>	Lys <sub>548</sub>
Tyr <sub>411</sub>	Glu <sub>551</sub>





E2F <sub>(409-426)</sub> residue	pRb residue
Tyr <sub>411</sub>	Ile <sub>532</sub>
Tyr <sub>411</sub>	Glu <sub>554</sub>
His <sub>412</sub>	Arg <sub>656</sub>
His <sub>412</sub>	Lys <sub>653</sub>
Gly <sub>414</sub>	Glu <sub>533</sub>
Gly <sub>414</sub>	Lys <sub>652</sub>
Leu <sub>415</sub>	Leu <sub>649</sub>
Leu <sub>415</sub>	Glu <sub>553</sub>
Leu <sub>415</sub>	Lys <sub>537</sub>
Glu <sub>417</sub>	Lys <sub>537</sub>
Gly <sub>418</sub>	Arg <sub>467</sub>
Glu <sub>419</sub>	Thr <sub>645</sub>
Arg <sub>422</sub>	Glu <sub>464</sub>
Asp <sub>423</sub>	Arg <sub>467</sub>
Leu <sub>424</sub>	Lys <sub>530</sub>
Phe <sub>425</sub>	Phe <sub>482</sub>
Phe <sub>425</sub>	Lys <sub>475</sub>

In a second aspect, the present invention provides a method to identify an agent which modulates the interaction between pRb and  $E2F_{(409-426)}$ , the method comprising:-

- a) combining together pRb, E2F<sub>(409-426)</sub> and an agent, under conditions in which pRb and E2F<sub>(409-426)</sub> form a complex;
  - b) growing a crystal of any pRb/E2F(409-426) complex; and
  - c) analysing the crystal structure to determine whether the agent is an agent which modulates the interaction between pRb and E2F<sub>(409-426)</sub>.



In the present invention, the term "modulates" is intended to refer to inhibiting, enhancing, destabilising and/or stabilising the interaction between pRb and E2F<sub>(409-426)</sub> and/or the formation of the pRb/E2F(409-426) complex and/or the stability of the complex after formation.

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"conditions in which pRb and E2F(409-426) can form a complex" are those conditions in which pRb and E2F<sub>(409-426)</sub> form a complex in the absence of an agent. Therefore the effect of the agent on the interaction between pRb and E2F(409-426) and complex formation can be assessed.

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Growing a crystal of a pRb/E2F<sub>(409-426)</sub> complex in step b) can be performed using methods well known to the person skilled in the art, for example using methods described in Practical Protein Crystallography 1999, McRee, D. E., Academic Press, San Diego, Ca, USA; and also in Protein Crystallization Techniques, Strategies and Tips 1999, Bergfors, T. M., International University Line, Ca, USA.

In the method, the combining of the pRb, E2F<sub>(409-426)</sub> and agent may be in any order. The order may be combining pRb with the agent and then adding the E2F<sub>(409-426)</sub>. Alternatively, the order may be combining E2F<sub>(409-426)</sub> with the agent and then adding pRb, or combining pRb with E2F<sub>(409-426)</sub> and then the agent. For example, the pRb may be combined with E2F<sub>(409-426)</sub> before soaking the complex in the agent, preferably in a solution of the agent. In this regard, two of the pRb, E2F(409-426) and agent may be cocrystalised before adding the pRb, E2F<sub>(409-426)</sub> or agent, as appropriate.

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Preferably step c) comprises comparing the crystal structure to the crystal structure of the first aspect of the invention.

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The agent may be selected using the three dimensional atomic co-ordinates of Annex



In a third aspect, the present invention provides a method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising selecting an agent using the threedimensional atomic coordinates of Annex 1.

Preferably, said selection is performed in conjunction with computer modeling. 5

Preferably the method comprises the further steps of:

- a) contacting the selected agent with pRb and E2F<sub>(409-426)</sub> under conditions in which pRb and E2F<sub>(409-426)</sub> can form a complex; and
- b) measuring the binding affinity of pRb to E2F<sub>(409-426)</sub> in the presence of the agent 10 and comparing the binding affinity to that of pRb to E2F(409-426) when in the absence of the agent, wherein an agent modulates a pRb/E2F(409-426) complex when there is a change in the binding affinity of pRb to E2F<sub>(409-426)</sub> when in the presence of the agent.

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## The method may further comprise:

- a) growing a supplementary crystal from a solution containing pRb and E2F<sub>(409-</sub> 426) and the selected agent where said agent changes the binding affinity of the pRb/E2F<sub>(409-426)</sub> complex under conditions in which pRb and E2F<sub>(409-426)</sub> can form a complex;
- b) determining the three-dimensional atomic co-ordinates of the supplementary crystal by X-ray diffraction using molecular replacement analysis;
- c) comparing the three dimensional atomic co-ordinates with those for the crystal structure as defined in the first aspect of the invention; and
- d) selecting a second generation agent using the three-dimensional atomic 25 coordinates determined for the supplementary crystal.

Preferably, said selection is performed in conjunction with computer modeling.





In a fourth aspect there is provided a method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:

- a) contacting a selected agent with pRb and E2F<sub>(409-426)</sub> under conditions in which pRb and E2F<sub>(409-426)</sub> can form a complex; and
- b) measuring the binding affinity of pRb to E2F<sub>(409-426)</sub> in the presence of the agent and comparing the binding affinity to that of pRb to E2F<sub>(409-426)</sub> when in the absence of the agent, wherein an agent modulates a pRb/E2F<sub>(409-426)</sub> complex when there is a change in the binding affinity of pRb to E2F<sub>(409-426)</sub> when in the presence of the agent.

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There is a "change in the binding affinity" when the binding affinity either decreases or increases when in the presence of the agent. If a decrease is observed, the agent may be inhibiting the complex. If an increase is observed, the agent may be enhancing the complex.

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The method of the fourth aspect may further comprise:

- a) growing a supplementary crystal from a solution containing pRb and E2F<sub>(409-426)</sub> and the selected agent where said agent changes the binding affinity of the pRb/E2F<sub>(409-426)</sub> complex under conditions in which pRb and E2F<sub>(409-426)</sub> can form a complex;
- b) determining the three-dimensional atomic coordinates of the supplementary crystal by X-ray diffraction using molecular replacement analysis;
- c) comparing the three dimensional atomic co-ordinates with those for the crystal structure defined in the first aspect of the invention; and
- d) selecting a second generation agent using the three-dimensional atomic coordinates determined for the supplementary crystal

Preferably, said selection is performed in conjunction with computer modeling.



In a fifth aspect, the present invention provides a method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:

- a) selecting an agent;
- b) co-crystalising pRb with the agent;
- c) determining the three dimensional coordinates of the pRb-agent association by X-5 ray diffraction using molecular replacement analysis; and
  - d) comparing the three dimensional coordinates with those of the crystal structure claimed in claim 1.
- In a sixth aspect, the present invention provides a method of identifying an agent that 10 modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:
  - a) selecting an agent;

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- b) crystalising pRb and soaking the agent into the crystal;
- c) determining the three dimensional coordinates of the pRb-agent association by Xray diffraction using molecular replacement analysis; and
  - d) comparing the three dimensional coordinates with those of the crystal structure claimed in claim 1.

In a seventh aspect, the present invention provides a method of identifying an agent 20 that modulates a pRb/E2F(409-426) complex, comprising:

- a) selecting an agent;
- b) co-crystalising pRb, E2F<sub>(409-426)</sub> and the agent;
- c) determining the three dimensional coordinates of the pRb-E2F-agent association by X-ray diffraction using molecular replacement analysis; and
- d) comparing the three dimensional coordinates with those of the crystal structure 25 claimed in claim 1.

In an eighth aspect, the present invention provides a method of identifying an agent that modulates a pRb/E2F(409-426) complex, comprising:

30 a) selecting an agent;

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- b) co-crystalising pRb and E2F<sub>(409-426)</sub> and soaking the agent into the crystal;
- c) determining the three dimensional coordinates of the pRb-E2F-agent association by X-ray diffraction using molecular replacement analysis; and
- d) comparing the three dimensional coordinates with those of the crystal structure claimed in claim 1.

Preferably the agent in the fifth, sixth, seventh or eighth aspect is selected using the three dimensional atomic co-ordinates of Annex 1. Preferably the method of the fifth, sixth, seventh or eighth aspect further comprises selecting a second generation agent using the three dimensional atomic coordinates determined. The second generation agent is preferably selected using the three dimensional atomic coordinates of Annex 1. The selection may be performed in conjunction with computer modeling.

Preferably the selected agent and/or the second generation agent, in the second, third, fourth, fifth, sixth, seventh and/or eighth aspects mimics a structural feature of  $E2F_{(409-426)}$  when said  $E2F_{(409-426)}$  is bound to pRb.

Preferably soaking refers to the pRb/E2F $_{(409-426)}$  complex being transferred to a solution containing the selected agent.

The method as defined in the third aspect preferably comprises the further steps of:

- a) contacting the selected agent with a pRb/E2F<sub>(409-426)</sub> complex; and
- b) determining whether the agent affects the stability of the complex.
- 25 Preferably the determination is with fluorescence polarization.

In a ninth aspect, the present invention provides a method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:

a) contacting a fluorescently tagged E2F<sub>(409-426)</sub> peptide (E2F-fluoropeptide) with pRb to allow pRb/E2F-fluoropeptide complex formation;





- b) detecting the fluorescence polarization;
- c) adding a selected agent; and
- d) detecting the fluorescence polarization in the presence of the agent.
- In a tenth aspect, the present invention provides a method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising;
  - a) contacting a fluorescently tagged E2F<sub>(409-426)</sub> peptide (E2F-fluoropeptide) with pRb to allow pRb/E2F-fluoropeptide complex formation;
  - b) detecting the fluorescence polarization;
- 10 c) contacting a selected agent with pRb and E2F<sub>(409-426)</sub> peptide (E2F-fluoropeptide) under conditions in which pRb and E2F-fluoropeptide can form a complex;
  - d) detecting the fluorescence polarization; and
  - e) comparing the fluorescence polarization detected in b) and d).
- Preferably the fluorescently tagged E2F peptide is selected using the three dimensional atomic co-ordinates of Annex 1.

Preferably a decrease in fluorescence polarization in the presence of the agent indicates that the agent destabilises the complex.

The methods of the ninth or tenth aspects may comprise the further step of adding untagged E2F<sub>(409-426)</sub> and detecting fluorescence polarization.

Preferably if fluorescence polarization decreases, on addition of the untagged E2F<sub>(409-426)</sub>, the agent does not stabilise the complex.

Preferably if there is no substantial change in fluorescence polarization, on addition of the untagged  $E2F_{(409-426)}$ , the agent stabilises the complex.

30 Preferably the method further comprises:





- a) contacting a fluorescently tagged E7 peptide (E7-fluoropeptide) with pRb to allow pRb/E7-fluoropeptide complex formation;
- b) detecting the fluorescence polarization;
- c) adding an agent that modulates pRb/E2F(409-426) complex; and
- 5 d) detecting the fluorescence polarization in the presence of the agent.

Alternatively the method may further comprise:

- a) contacting a fluorescently tagged E7 peptide (E7-fluoropeptide) with pRb to allow pRb/E7-fluoropeptide complex formation;
- 10 b) detecting the fluorescence polarization;
  - c) contacting an agent that modulates pRb/E2F<sub>(409-426)</sub> complex with pRb and E7-fluoropeptide under conditions in which pRb and E7-fluoropeptide can from a complex;
  - d) detecting the fluorescence polarization; and
- e) comparing the fluorescence polarization detected in b) and d).

Preferably a decrease in fluorescence polarization indicates that the agent also inhibits E7 binding to pRb. Such agents can then be removed from the method because the agents are identified as non-specific inhibitors. This identification of non-specific inhibitors can dramatically reduce the workload downstream of the assay method, for example in biochemical assays, thereby accelerating the hit to lead discovery process.

In addition ANS (aniline naphthalene sulphonic acid) reagent may be used to detect hydrophobic surfaces on pRb which become exposed as it unfolds. The fluorescence of ANS is highly sensitive to its environment. In solution there is virtually no fluorescence, whereas when bound to protein, such as pRb, it fluoresces highly. Changes in protein structure can alter the fluorescent signal of bound ANS due to changes in its environment to water. Therefore changes in pRb structure can be detected on addition of ANS and the agent that modulates pRb/E2F<sub>(409-426)</sub> complex. If the fluorescent signal alters on addition of the agent, the agent may be affecting the pRb



structure. The use of ANS to monitor protein unfolding is known in the art (Semisotnov et al (1991) Biopolymers, 31(1), 119-128)

The binding affinities may be measured by isothermal titration calorimetry.

5 Alternatively the binding affinities may be measured by Surface Plasmon Resonance (SPR).

In an eleventh aspect, the present invention provides an agent identified by a method according to the second, third, fourth, fifth, sixth, seventh, eighth, ninth and/or tenth aspects of the invention.

In a twelfth aspect, the present invention provides an agent, as set out according to the eleventh aspect of the invention, for use as an apoptosis promoting factor in the prevention or treatment of proliferative diseases.

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Preferably the, or each selected agent is obtained from commercial sources or is synthesised. Preferably the agent is for use in preventing or treating cancer, which may be pancreatic cancer and related diseases.

In a thirteenth aspect, the present invention provides the use of an agent, which modulates a pRb/E2F<sub>(409-426)</sub> complex, identified by a method according to the second, third, fourth, fifth, sixth, seventh, eighth, ninth and/or tenth aspects of the present invention, in the manufacture of a medicament for the prevention or treatment of proliferative diseases.

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The proliferative diseases may be cancer, preferably pancreatic cancer and related diseases.





In a fourteenth aspect, the present invention provides the use of the atomic coordinates of the crystal structure as set out according to the first aspect of the present invention, for identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex.

- In a fifteenth aspect, the present invention provides computer readable media comprising a data storage material encoded with computer readable data, wherein said computer readable data comprises a set of atomic co-ordinates of the pRb/E2F<sub>(409-426)</sub> crystal structure according to Annex 1 recorded thereon.
- Preferred features of each aspect of the invention are as for each of the other aspects mutatis mutandis.

The present invention will now be described, by way of example only, and with reference to the following figures, in which:

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Annex I.

Atomic co-ordinates for crystal of pRb/E2F<sub>(409-426)</sub> complex.

In Annex 1 there is shown:

Column Number	Description	
2	Atom number	
3	Atom type	
4	Residue type	
5	pRb domains (A or B) or E2F <sub>(409-426)</sub> (P)	
6	Residue number	
7	x co-ordinate of atom (Å)	
8	y co-ordinate of atom (Å)	
9	z co-ordinate of atom (Å)	
10	Occupancy	
11	B-factor (Å <sup>2</sup> )	

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## Figure 1.

Structure of pRb/E2F.

- (A) Schematic drawing of functional domains and protein constructs used for pRb, E2F. The shading used for the constructs in this panel match those used in subsequent figures.
- (B) The structure of  $pRb_{AB}/E2F_{(409-426)}$ , shown in two orthogonal views in Ribbons representation. The helices of the A domain are shown as a darker shade to those of the B domain. The main-chain trace of E2F is labelled.

(C) The interactions between E2F<sub>(409-426)</sub> and pRb<sub>AB</sub> are shown schematically with the E2F peptide running down the centre. Residues of E2F that are conserved across the five family members are shown as ovals, while the five residue subset of these conserved residues whose mutation leads to disruption of the pRb/E2F interaction are shaded. Hydrogen-bond interactions are shown as broken lines, while hydrophobic contacts are indicated by bands. Residues from domain A of pRb are labelled with an asterisk and the other residues are from domain B. All of the pRb residues shown are either invariant or conserved across 27 species of pRb, p107 and p130.

## 20 Figure 2.

Isothermal Titration Calorimetry (ITC) measurements.

- (A) The upper panel shows the raw data of an ITC experiment performed at  $22^{\circ}$ C. The lower panel shows the integrated heat changes, corrected for the heat of dilution, and the fitted curve based on a single site model. The panel represents the experiment where  $E2F_{(243-437)}$  is titrated into Rb<sub>AB</sub>.
- (B) Summary of dissociation constants obtained by ITC measurements. The quoted errors are those produced by fitting the data to a two-state model. For the interaction of E2F<sub>(243-437)</sub> to Rb<sub>AB</sub> and Rb<sub>ABC</sub> the affinities are too high to measure reliably and we have therefore quoted the upper limits of the dissociation constants.

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Figure 3 - Binding of Fluorescein-E2F, Rhodamine-E2F and Fluorescein-E7 to pRb

Figure 4 - Displacement binding curves: a) E2F<sub>409-426</sub> peptide; b) detergent

Figure 5 – Screen controls from test screen 6 x 384 plates

- Figure 6 Correlation between inhibition of Rhodamine and Fluorescein-E2F
- 10 Figure 7 Correlation between inhibition of Fluorescein-E2F and Fluorescein-E7
  - Figure 8 a) Titration curves of rho-N-E2F (n=3); b) Time course of the change of fluorescence polarization signal with time taken from a test screen (mean±s.e.m., n=960)

Figure 9 – IC50 curves determined for hits identified using the screening protocol described with reference to Figures 3 to 8: a) hit compound IC50 curve; b) non-specific inhibitor IC50 curve

## 20 Structure determination of pRb/E2F

For crystallisation we used a pRb construct based on that previously described by Lee, J.O., Russo, A.A., and Pavletich, N.P. (1998). Structure of the retinoblastoma tumour-suppressor pocket domain bound to a peptide from HPV E7, Nature 391, 859-65, which has engineered thrombin cleavage sites at the ends of the flexible linker region between the A and B domains. Purification and proteolysis produces a final protein containing residues 372 to 589 of the A domain and 636 to 787 of the B domain (hereafter pRb<sub>AB</sub> – Figure 1A). Although these two domains are not covalently attached after thrombin treatment, they remain tightly associated. The removal of the A-B linker region facilitates crystallisation of pRb but does not alter its binding affinity for E2F. Crystals of the pRb/E2F<sub>(409-426)</sub> complex grew in a plate-like

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habit with typical dimensions 200 x 200 x 10  $\mu m^3$ . Repeated attempts at data collection from flash-cooled crystals using synchrotron X-ray sources were thwarted by very high crystal mosaicity and poor data reduction statistics. The problem was overcome by using the micro-focus diffractometer on station ID13 at ESRF current experience and plans at EMBL and ESRF/ID13, Acta Crystallogr D 55, 1765-1770), currently the only such device installed at a synchrotron source. Using a  $10x10 \ \mu m^2$ aperture, data were collected from four separate and widely spaced volumes of a single crystal in order to minimise radiation damage. A total of 100, 1° oscillation images were collected using a MAR CCD detector. These data extended to a Bragg spacing of 2.5 Å with an overall  $R_{merge} = 9.2\%$ , and completeness of 87%. The structure was solved by molecular replacement and produced initial electron density maps in which the E2F peptide (E2F<sub>(409-426)</sub>) could be readily located.

#### Protein constructs.

Rb<sub>AB</sub> was expressed as a GST-fusion protein in E. coli using the pGEX-6P vector. 15 The construct was engineered to contain a Prescission protease site at the N-terminus of Rb as well as two thrombin sites (LVPRGS) inserted at either end of the flexible A-B linker. Fusion protein was loaded onto a glutathione Sepharose 4B column before treatment with thrombin and Prescission protease. The resulting eluent was further purified using a Superdex 200 gel filtration column. RbABC was expressed in 20 E. coli with a C-terminal His-tag using pET-24. Crude lysate was first purified using an S-sepharose column followed by a Ni-NTA step before being run on a Superdex 200 gel filtration column. Recombinant human E2F1(243-437) was expressed in E. coli using pGEX-6P with an engineered Prescission protease site at the N-terminus of E2F. Crude lysate was bound onto a glutathione Sepharose 4B column prior to cleavage 25 with the protease. The resulting eluent was further purified by gel filtration on a Superdex 75 column. E2F<sub>(409-426)</sub> and E2F<sub>(380-437)</sub> were synthetic peptides. HPV-16 E7(17-98) was prepared as described elsewhere (Clements, A.J., K, Mazzareli, J.M. Ricciardi, R.P. Marmorstein R. (2000). Oligomerization properties of the viral



oncoproteins adenovirus E1A and human papillomavirus E7 and their complexes with the retinoblastoma protein., Biochemistry 39, 16033-16045).

## Crystallography.

Plate-like crystals were grown by the hanging drop vapour diffusion method at 4°C. 5 Rb<sub>AB</sub> was mixed with the E2F-1 peptide at 1:2 molar ratio and concentrated to 15mg/ml. Hanging drops were formed by mixing 1µl of protein solution with an equal volume of reservoir solution containing; 0.14M Na citrate, 26% PEG400, 4% n-propanol and 0.1M Tris at pH 7.8. Crystals were flash frozen in mother-liquor made up to 25% glycerol. Diffraction data were collected using the micro-focus 10 diffractometer at ESRF and processed using the DENZO and SCALEPACK software (Otwinowski, Z.M., W. (1993). In Data Collection and Processing, L.I. Sawyer, N. Bailey, S., ed. (SERC Daresbury Laboratory), pp. 556-562). Molecular replacement calculations were carried out using Amore (CCP4, 1994) with 1GUX.brk as the search model. The final model contains co-ordinates for the protein which cover residues 15 379-578 of the A domain and 644-787 of the B domain of Rb and the entire E2F<sub>(409-</sub> 426) peptide for the four copies present in the asymmetric unit together with 600 solvent molecules. The refined model has the following residuals;  $R_{work} = 23.7\%$ ,  $R_{free}$ =28.7%, rmsd bonds = 0.007 Å, rmsd angles =1.3°.

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## Structure of pRb/E2F complex

The packing of the A and B domains generates a waist-like interface groove into which E2F<sub>(409-426)</sub> binds in a largely extended manner (Figure 1B). The peptide makes contacts with residues from helices  $\alpha 4$ ,  $\alpha 5$ ,  $\alpha 6$ ,  $\alpha 8$  and  $\alpha 9$  of domain A, and with  $\alpha 11$ from domain B of pRb. Formation of the complex buries 2280 Å<sup>2</sup> of surface area, 1500  $\text{Å}^2$  of which are hydrophobic. The two end regions of the E2F<sub>(409-426)</sub> fragment make extensive contacts with pRb, while interactions made by the middle section of the E2F<sub>(409-426)</sub> fragment (residues 416 to 420) are relatively sparse (Figure 1C). Overall, a high proportion of the hydrogen bond interactions between the two

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molecules involves the side chains of conserved pRb residues interacting with the main chain of E2F. Examination of the distribution of conserved residues over the surface of pRb, reveals that the majority are accounted for by the E2F binding site. There is a somewhat smaller cluster of conserved residues associated with the LxCxE binding site. Perhaps the most remarkable aspect of this analysis is that although pRb has been reported to associate with at least 110 cellular proteins perhaps 50 or more in a pocket-dependent manner, the E2F and LxCxE binding sites account for almost all of the conserved residues on its surface. There are two explanations that may partially account for these observations. Firstly, many of the reported binding partners of pRb have yet to be verified. Secondly, the LxCxE binding site is probably responsible for mediating the binding of many different proteins, such as HDAC, to pRb.

Since there are four copies of the pRb/E2F<sub>(409-426)</sub> complex in the asymmetric unit of our crystal form it is possible both to compare these four crystallographically independent copies of the pRb/E2F(409-426) complex and to compare them with the crystal structure of pRb/E7 without bond E2F (Lee et al., 1998 Supra). The first six residues at the N-terminus, the  $\alpha 3-\alpha 4$  and  $\alpha 6-\alpha 7$  loops adopt different conformations between the four copies in our asymmetric unit, while the variations across the rest of the structure between the four molecules is not significant. Comparison of the pRb structure in the presence and absence of bound E2F<sub>(409-426)</sub> shows that there is essentially no change in the relative orientation of the two lobes of the A/B pocket on E2F<sub>(409-426)</sub> binding nor any widespread changes in the structures of the individual domains. This comparison does reveal that the end of  $\alpha 4$  and the connecting loop to α5 becomes ordered in the pRb/E2F<sub>(409-426)</sub> complex as two conserved residues (Glu464-pRb & Arg467-pRb located towards the end of  $\alpha 4$  in our structure) interact with the E2F<sub>(409-426)</sub> peptide. Within the E2F<sub>(409-426)</sub> construct there are eight residues that are conserved across E2F's from all animal species (Figure 1A). Amino-acid substitutions at five of these positions have been shown to lead to loss of binding to pRb but retention of E2F's trans-activation potential. The following description



focuses on the structural role of these five residues. Tyr(411)-E2F appears to play an important role in peptide binding because its phenolic ring occupies a hydrophobic pocket created by Ile(536)-pRb, Ile(532)-pRb, Ile(547)-pRb and Phe(413)-E2F, while its hydroxyl group makes a hydrogen bond to the invariant Glu(554)-pRb. Towards the C-terminal part of the E2F peptide, Leu(424)-E2F and Phe(425)-E2F make several hydrophobic interactions, two of which involve conserved residues. Leu(424)-E2F makes contacts with the aliphatic portion of the side chain of Lys(530)-pRb and also packs against Leu(415)-E2F and Phe(425)-E2F. In addition, Phe(425)-E2F itself packs against Phe(482)-pRb. Unlike the residues of E2F just discussed, the side-chains of Glu(419)-E2F and Asp(423)-E2F do not point into the groove formed between the A and B domains of pRb, but instead point away from it. Glu(419)-E2F hydrogen bonds through a water molecule with the main-chain carbonyl of Thr(645)-pRb; Asp(423)-E2F forms a salt bridge with Arg(467)-pRb located at the end α4.

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Finally, as described earlier, the crystal structure shows how E2F makes extensive contacts with largely conserved residues from both the A and B domains of the pocket and that the binding site for E2F is dependent on the structure of the interface between the two domains. This feature of the structure suggests that E2F acts as a sensor of the structural integrity of the pRb pocket. The position and nature of the E2F binding site make the binding of the transcription factor particularly sensitive to mutations in the pocket region of the tumour suppressor protein. The potential significance of these observations will be discussed later with regard to the normal role of pRb in protecting cells against E2F-mediated apoptosis.

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## Additional determinants of pRb/E2F function

It is clear from a number of studies that, although E2F<sub>(409-426)</sub> expressed as a Gal4 fusion protein is sufficient to recruit pRb and repress transcription, there are additional interactions made by the physiologically relevant E2F/DP heterodimer with pRb.

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Similarly, while the pocket domain is highly conserved, the most frequent site of deleterious mutation, and capable of transcriptional repression, it is not sufficient for the physiological function of pRb. In particular, the C-terminus of pRb is necessary for mediating growth arrest and recruitment of certain cyclin/cdk complexes as well as containing several of the residues whose phosphorylation leads to deactivation of pRb function. Therefore, in order to better understand the requirements for physiological pRb/E2F complex formation, we have made a series of constructs of the two proteins (Figure 1A) and carried out binding measurements by isothermal titration calorimetry (ITC). We have also carried out a series of competition experiments to confirm qualitatively the interpretation of the ITC binding data.

## Isothermal Titration Calorimetry.

Binding of the various E2F constructs to  $Rb_{AB}$  and  $Rb_{ABC}$  was measured by isothermal titration calorimetry using a MicroCal Omega VP-ITC machine (MicroCal Inc., Northampton, USA). The E2F constructs at a concentration between 100-150  $\mu M$ 

were titrated into 12-15 μM Rb at a temperature of 22°C. Proteins were dialysed against 50mM Tris pH 7.6, 100mM NaCl and 1mM TCEP. After subtraction of the dilution heats, calorimetric data was analysed using the evaluation software MicroCal Origin v5.0 (MicroCal Software Inc.). For all of the titrations, the stoichiometry of ligand binding to Rb was very close to 1.0. For E2F<sub>(243-437)</sub> binding to Rb, the binding affinity and the heat change associated with binding were such that we could only establish that binding was tighter than 10 nM. In order to verify that binding of this protein was similar for both Rb<sub>AB</sub> and Rb<sub>ABC</sub> we carried out competition experiments

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## Competition experiments.

The proteins used in these experiments were His<sub>6</sub>-Rb<sub>ABC</sub> (RESIDUES 380-929); MW 66.07kDa, non-tagged Rb<sub>AB</sub> (residues 372-787); MW 48.67 KDa, are His<sub>6</sub>-Rb<sub>AB</sub> (residues 376-792); MW 49.86 KDa, E2F<sub>(243-437)</sub>; MW 21.45 KDa HPV E7 (residues

which showed approximately equal partition between the two different Rb proteins.



17-98); MW 9.38 KDa and E2F<sub>(409-426)</sub>; MW 2.12 KDa. Protein concentrations were carefully determined by u.v. spectroscopy and confirmed by ITC titrations. The small acidic E2F proteins stain much weaker than Rb with Coomassie on SDS-PAGE. For all gel lanes contained a final Rb<sub>AB</sub> concentration of ca. 7μM. After equilibration with E2F<sub>(243-437)</sub> and E2F<sub>(409-426)</sub> the samples were loaded onto a 1.0ml Ni column and washed with 7 x 0.5 ml of loading buffer (50mM Tris pH 7.5, 200mM NaCl & 10mM Imidazole). The samples were then eluted with 7 x 0.5ml elution buffer (50mM Tris, pH 7.5, 200mM NaCl, 200mM Imidazole). After volume correction samples were boiled in SDS loading buffer and run on a 4-12% SDS PAGE. For the two pRb proteins and E2F<sub>(243-437)</sub> were mixed at 40μM in a final volume of 0.5ml. After equilibration for 2hrs the mixture was loaded onto 1ml Ni beads in a small column, washed with 7 x 0.5ml of loading buffer (50mM Tris, pH 7.5, 200mM NaCl, 10mM Imidazole), eluted using 7 x 0.5ml elution buffer (50mM Tris, pH 7.5, 200mM NaCl, 200mM Imidazole). Samples, after correcting for volume were boiled in SDS sample buffer and run on a 4-12% SDS gel.

A typical ITC experiment is shown in Figure 2A and a summary of the affinity constants obtained for both pRb<sub>AB</sub> and pRb<sub>ABC</sub> interacting with three constructs of E2F are given in Figure 2B. The two shorter E2F constructs bind to either pRb<sub>AB</sub> or pRb<sub>ABC</sub> with similar affinities. However, E2F<sub>(243-437)</sub> binds at least 16-fold stronger than either of the two shorter E2F fragments to both pRb<sub>AB</sub> and Rb<sub>ABC</sub>. Our ITC data therefore show that there are additional interactions of the A/B pocket of pRb with a region of E2F-1 outside of the transactivation domain. This result has been confirmed qualitatively by competition experiments which show that a 15-to 30-fold molar excess of the shorter E2F peptide is required to 50% compete with E2F<sub>(243-437)</sub> for binding to pRb. Our results are consistent with an earlier report that noted an interaction of pRb with the marked box region of E2F (residues 245-317). Taken together, these data demonstrate that the majority of the free energy of interaction between pRb and E2F is contributed by the 18-residue segment E2F<sub>(409-426)</sub> used in our



structure analysis. There is an additional stabilising interaction between the marked box region of E2F and pRb, that contributes approximately 2kcal mol<sup>-1</sup> to the overall free energy of complex formation, but is not sufficient on its own for complex formation.

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As the binding constant for the interaction of E2F<sub>(243-437)</sub> with pRb<sub>AB</sub> (or pRb<sub>ABC</sub>) was too tight to determine reliably by ITC we performed a competition experiment to determine if this E2F construct bound preferentially to one or the other pRb construct. The results show approximately equal partitioning of E2F(243-437) between the two pRb species and demonstrates therefore, that the C-terminus of pRb does not participate in the binding to E2F-1 in isolation. This means that in addition to the interaction of E2F<sub>(409-426)</sub> with the pocket region of pRb there is an additional interaction, almost certainly involving the marked box region of E2F, that also binds to the pRb pocket. This data is consistent with the hypothesis that the approximately 10-fold stronger interaction of E2F/DP with pRbABC rather than pRbAB reported previously arises through interactions of the DP component of the E2F/DP heterodimer with the C-terminus of pRb. This idea is strongly supported by the data from another study which shows that DP-1 interacts with pRb in a manner that does not require the structural integrity of the A/B pocket. Taken together, these data indicate that at least one of the functions of the C-terminus of pRb is to bring additional stabilisation to the interaction of the tumour suppressor with the heterodimeric E2F/DP transcription factors.

## Use of structure atomic co-ordinates of Annex I

The atomic co-ordinates of Annex 1 are cartesian co-ordinates derived from the results 25 obtained on diffraction of a monochromatic beam of X-rays by the atoms of the pRb/ E2F<sub>(409-26)</sub> complex in crystal form. The diffraction data was used to calculate electron density maps of the crystal. The electron density maps were then used to position the individual atoms of the pRb/ E2F(409-26) complex.

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The determination of the three-dimensional structure of the pRb/E2F<sub>(409-426)</sub> complex provides basis for the design of new and specific agents that modulates formation of the complex and/or modulates the interaction between pRb and E2F<sub>(409-426)</sub>. For example, computer modelling programs may be used to design different molecules expected to modulate formation of the pRb/E2F<sub>(409-426)</sub> complex and/or the interactions between pRb and E2F<sub>(409-426)</sub>.

A candidate agent, may be any available compound. A commercial library of compound structures such as the Cambridge Structural Database would enable computer based *in silico* screening of the databases to enable compounds that may interact with, and/or modulate formation of, the complex to be identified.

Such libraries may be used to allow computer-based high throughput screening of many compounds in order to identify and select those agents with potential to modulate formation of the  $pRb/E2F_{(409-426)}$  complex and/or the interaction between pRb and  $E2F_{(409-426)}$ .

In this regard, a potential modulating agent can be subjected to computer modelling with a docking program such as GRAM, DOCK or AUTODOCK (see Walters et al., Drug discovery Today, Vol.3, No. 4, (1998), 160-178, and Dunbrack et al., Folding and Design, 2 (1997) 27-42) to identify and select potential agents. This can include computer fitting of potential modulating agents to the pRb/E2F<sub>(409-426)</sub> complex to ascertain how the agent, in terms of shape and structure, will bind to the complex.

Computer programs can be employed to estimate the interactions between the pRb, E2F<sub>(409-426)</sub> and agent or pRb/E2F<sub>(409-426)</sub> complex and agent. These interactions may be attraction, repulsion, and steric hindrance of the two binding partners (e.g. the pRb/E2F<sub>(409-426)</sub> complex and a selected agent). A potential agent will be expected to be more potent if there is a tighter fit and fewer steric hindrances, and therefore greater attractive forces. It is advantageous for the agent to be specific to reduce interaction

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with other proteins. This could reduce the occurrence of side-effects due to additional interactions with other proteins.

Potential agents that have been designed or selected possible agents can then be screened for activity as set out in the second to tenth aspects above. The agents can be obtained from commercial sources or synthesised. The agent is then contacted with  $pRb/E2F_{(409-426)}$  complex to determine the ability of the potential agent to modulate the formation of the complex. Alternatively the agent may be contacted with pRb and  $E2F_{(409-426)}$  under conditions in which pRb and  $E2F_{(409-426)}$  can form a complex (in the absence of agent), to determine the ability of the agent to modulate complex formation.

A complex of pRb/E2F<sub>(409-426)</sub> and said potential agent can then be formed such that the complex can be analysed by X-ray crystallography to determine the ability of the agent to modulate complex formation and/or the interaction between pRb and E2F<sub>(409-426)</sub>.

The complex of pRb/E2F $_{(409-426)}$  and agent could be compared to that for pRb/E2F $_{(409-426)}$  alone with the three dimensional atomic co-ordinates in Annex 1.

Detailed structural information can then be obtained about the binding of the potential agent to the complex,. This will enable the structure or functionality of the potential agent to be altered to thereby to improve binding. The above steps may be repeated as may be required.

The agent-pRb/E2F<sub>(409-426)</sub> complex could be analysed by co-crystallising pRb/E2F<sub>(409-426)</sub> with the selected agent or soaking the agent into crystals of the pRb/E2F<sub>(409-426)</sub> complex; and then determining the three dimensional co-ordinates of the agent-complex by X-ray diffraction using molecular replacement analysis.

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Therefore, the pRb/E2F<sub>(409-426)</sub> -agent complexes can be crystallised and analysed using X-ray diffraction data obtained and processed, for example using the DENZO and SCALEPACK software (Otwinowksi, Z. M., W. (1993). Difference Fourier electron density maps can be calculated based on X-ray diffraction patterns of soaked or co-crystallised pRb/E2F<sub>(409-426)</sub> complex and the solved structure of uncomplexed agent. These maps can then be used to determine the structure of the agent bound to the pRb/E2F<sub>(409-426)</sub> and/or changes in the conformation of pRb/E2F<sub>(409-426)</sub> complex relative to the pRb/E2F<sub>(409-426)</sub> complex in the absence of agent.

- The agent may be improved, for example by adding or removing functional groups, substituting groups or altering its shape in light of data obtained from agent bound to pRb/E2F<sub>(409-426)</sub> complex and/or agent bound to pRb. Such an improved agent may then be subjected to the methods of the invention.
- 15 Electron density maps can be calculated using programs such Amore from the CCP4 computing package (Collaborative Computational Project 4. The CCP4 Suite: Programs for Protein Crystallography, Acta Crystallographical, D50, (1994), 760-763).
- The provision of computer readable media enables the atomic co-ordinates to be accessed to model the pRb/E2F<sub>(409-426)</sub> complex by, for example, RAMSOL (a publicly available computer software package which allows access and analysis of atomic co-ordinate data for structure determination and/or rational drug design).
- In addition, structure factor data, derivable from the atomic co-ordinate data (see e.g. Blundell et al., in Protein Crystallography, Academic Press, New York, London and San Francisco, (1976)), can be used to enable difference Fourier electron density maps to be deduced.

## 30 Screening assays



After an agent has been selected, its inhibitory effect on pRb/E2F(409-426) complex formation or ability to interact with the pRb/E2F(409-426) complex can be assessed with one or more of the methods of the invention.

For example, the crystal structure of the interaction of E2F<sub>(409-426)</sub> with pRb can be 5 used to aid the design of a fluorescently tagged peptide for the use in a binding assay suitable for high throughput screening of low molecular weight compounds or peptide libraries. The fluorescent tag may be a fluorescein, rhodamine or some other commercially available tag chemically attached via a suitable amine or thiol group.

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Binding could be measured by detecting fluorescence polarization in an homogeneous assay format (i.e. one in which all reagents are mixed in a single well, and reaction occurs in solution without wash steps). Fluorescence polarization technology is commonly applied in high throughput screening laboratories (ref: Sokham et al. (1999) Analytical Biochemistry, 275, 156-161. "Analysis of protein-peptide interaction by a miniaturised fluorescence polarization assay using cyclin-dependent kinase2/cyclin E as a model system.")

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Fluorescence polarization can be used to determine binding of a fluorescently-tagged small molecule (ligand or peptide) with a large molecule (receptor or protein) by detecting changes in the rotational velocity of the small molecule in the free and bound state. The rotational velocity is inversely proportional to the size of the molecule. Using a fluorescently tagged peptide and suitable optics the changes in rotational velocity upon binding to pRb can be measured as a differences in light emitted in parallel and perpendicular to a polarized excitation source. Fluorescence polarisation gives a measure of the proportion of fluorescently tagged peptide found in the bound state in a homogenous format.

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In an assay method of the present invention, fluoro-peptide (E2F<sub>(409-426)</sub>fluoropeptide) bound to pRb will have a low rotational velocity and will appear





stationary during the excitation period. Emitted light will be transmitted in parallel to the polarized incident light and the light detected will have a high polarization value. In contrast in the presence of an inhibitor of the interaction between pRb and E2F<sub>(409-</sub> 426) -fluoropeptide, the free E2F<sub>(409-426)</sub> - fluoro-peptide will have a high rotational velocity and light will be transmitted in all directions. Emitted light will be detected both parallel and perpendicular to the polarized excitation source, and will have a low polarization value.

An example of the use of fluorescence polarisation is now described.

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Data from a Fluorescence Polarisation (FP) screen configured for the interaction of pRb with E2F is presented. Fluorescein-tagged E2F peptide was used to screen 10,000 small drug like molecules. Hit confirmation strategies based on fluorescence interference and specificity were developed and compared.

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Based on the crystal structure defined by the atomic co-ordinates in Annex 1, an FP screen was configured for the interaction of recombinant pRb A/B domains with E2F(409-426) peptide (see Fig 1B). In addition, a second peptide binding site (E7, see Fig 1B), distant from the E2F binding pocket, was utilised as an internal control for non-specific inhibitors. Fluorophores in the form of fluorescein and rhodamine labelled peptides were synthesised and were used in a primary screen and hit confirmation.

Knowledge of the interaction of E2F and E7 peptides with pRb influenced the design of the fluoro-peptides used in the assay. The following peptides were synthesised, labelled and tested.

N-terminal amide linkage 5carboxyfluorescein-E2F409-426, 18'mer. 1. (fl-N-E2F18)



#### LDYHFGLEEGEGIRDLFD

Rhodamine label at C-terminal cysteine E2F409-427, 19'mer. (Rh-C-2. E2F19)

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#### LDYHFGLEEGEGIRDLFDC

3. Rhodamine label at DDC substitution E2F409-426, 18'mer. (Rh-N2-E2F18)

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## LCYHFGLEEGEGIRDLFD

4. N-terminal amide linkage 5carboxyfluorescein-E7, nonomer (Fl-E7)

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## DLYCYEQLN

Peptides 1, 3 and 4 were used in the screen and subsequent hit confirmation assays.

Synthetic peptides were synthesised and fluoro-tagged using either N-terminal labelling with 5 carboxyfluorescein succinimidyl ester or cysteine labelling with single 20 isomer tetramethylrhodamine-5- maleimide. Typical titration binding curves of pRb with the fluoro-labelled peptides are shown (mean±sem, n=3) in Figure 3. Fluorescein fluorescence measured at  $\lambda$ excite = 485 and  $\lambda$ emit = 520 nm Rhodamine fluorescence measured at  $\lambda$ excite = 545 and  $\lambda$ emit = 580 nm

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Measurements were made using BMG Fluorostar plate reader fitted with polarization optic. Fluorescein-E2F showed the greatest degree of polarization, and consequently the best signal to noise. It was chosen as the label of choice for a primary screen. Data were fitted to a one site binding model using Graphpad prism. Kd values of



450± 70 and 380±50 nM were calculated for fluorescein and Rhodamine labelled E2F, which were similar to Kd determined for unlabelled peptide using isothermal calorimetry. Fluorescein-E7 showed tightest binding with Kd= 130±20 nM.

5 The assay principle was validated using unlabelled E2F peptide to displace Fl-E2F without disrupting Fl-E7 binding to pRb. Fluoro-tagged peptide (400 nM) was preincubated with pRb (1 μM) and unlabelled peptide added at the concentrations shown. Displacement binding curves were plotted (figure 4a), and were fitted to a one site competition binding model using Graphpad prism curve fitting software. These curves were compared to the effects of a detergent-like compound (figure 4b), which causes gross structural changes and disrupts binding of both peptides.

The results show that labelled E2F (F1-E2F) does not displace E7, thereby validating the assay principle.

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The assay was optimised in 384-well black plates (Matrix) and automated using a Beckman Fx liquid handling robot. 1  $\mu$ M pRb in 50 mM Tris HCL, pH7.0, 100 mM NaCl, 10 mM DTT, 0.05% NP-40 was mixed with 40  $\mu$ M compound (4% DMSO) and 0.4  $\mu$ M fluorescein-E2F (final concentrations). Controls from a test screen of 10,000 compounds are shown in Figure 5.

Polarized and depolarized signal from fluorescein-E2F with and without pRb present are shown in Figure 5 (solid and open circles respectively). Specific disruption of binding by E2F protein and peptide are also shown. Addition of E2F protein completely displaces F1-E2F (open triangle) and the signal is reduced to that of free fluoro-peptide alone. Addition of unlabelled-E2F at a concentration which gave 50% inhibition is clearly separated from the control populations. Hits were identified as compounds which reduced the polarization signal to less than mean-3sd of the fluorescein-E2F: pRb control.





## **Summary of Screen Data**

Assay Principle	Fluorescence Polarization
Assay Automation	Biomek Fx
Assay Detection	BMG Polar Star Reader
Assay Parameters Signal: noise	6.9
Signal: background	4.8
Z'	0.67
Test Screen 10,000 Z	0.45
Hit rate	0.93%

5 Z factors are statistical factors well known by the skilled person in the art. The Z' factor is defined by

$$Z' = 1 - {\frac{(3X \text{ s.d.of positive control} + 3X \text{ s.d of negative control})}{(\text{mean of positive control} - \text{mean of negative control})}}$$

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In the present assay:-

positive control = fully polarized signal; pRb plus fluoro-tagged E2F peptide negative control = depolarized signal from fluoro-tagged E2F peptide alone.

15 Z is calculated in much the same way except:

Positive control = polarized signal of pRb and fluoro-tagged E2F in presence of compounds.

## HIT CONFIRMATION:

20 Identification of Fluorescence Interfering Compounds.



A large proportion (37.5%) of the hits selected from the primary screen were coloured compounds which significantly altered the fluorescence intensity signal, and were potentially interfering with the assay. All hits were included in hit confirmation assays.

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Hits were re-plated from master stocks and re-tested against fluorescein-E2F and rhodamine-E2F. A correlation ( $r^2$ =0.69) between inhibition of fluorescein E2F and Rhodamine-E2F was observed (figure 6) with a hit confirmation rate of 78%. Notably, 60% of compounds which were potentially interfering with the fluorescein signal were inhibitors with Rhodamine-E2F assay, without affecting rhodamine fluorescence intensity signal. Suggesting that deselection of compounds on the basis of fluorescence interference can lead to loss of real inhibitors.

Finally the hits were tested against a second peptide binding site. Fluorescein-E7 peptide at 400 nM. The results were compared to inhibition of E2F and a scatter plot is shown in Figure 7. A weak correlation was observed (r<sup>2</sup>=0.51), with 72% of the inhibitors of E2F also inhibiting fluorescein E7. These compounds were excluded as non-specific inhibitors and were not taken forward in subsequent biochemical assays.

20 Comparison of Hit Confirmation Strategies on 80 best hits selected from a Primary screen of 10,000 compounds.

**Hit Confirmation rates** 

Confirmation Test	% Primary Hits
1. Inhibition in retest Fluorescein-E2F	77.5
2. Fluorescence Interference	37.5
3. Inhibition in retest Rhodamine-E2F	62.5
4. Inhibition of Fluorescein-E7	58.5

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# The impact of selection strategy on number of compounds selected for further biochemical study (eg IC50, isothermal calorimetry, co-crystallisation)

Strategy 1	Strategy 2	Strategy 3
Tests 1+2 Remove fluorescence interfering compounds	Tests 1+3 Select inhibitors active for both fluorescein- and rhodamine-E2F	Tests 1+3+4 E2F inhibitors but not E7 inhibitors
36	50	14
False Negatives	False Positives	Specific Compounds only

To demonstrate the stability and rapidity of binding equilibria of fluoro-peptide with pRb. The titration curves shown in Figures 8a and 8b are typical of several experiments and are of rho-N-E2F (n=3). The time course shown of the change of fluorescence polarization signal with time is taken from a test screen (mean ± s.e.m., n= 960).

pRb titration curves were performed in 96-well black plates, in a total reaction volume of 100uL. Doubling dilutions from 10  $\mu$ M stock of pRb were made in binding buffer (50 mM Tris HCL, pH7.0, 100 mM NaCl, 10 mM DTT, 0.05% NP-40) and 80  $\mu$ L added in triplicate to wells. 20  $\mu$ L of 2  $\mu$ M fluoro-peptide was added and pipetted up and down to mix. The plate was read after 1 hr incubation at room temperature.

Compound interference was not a useful factor upon which to deselect compounds in an FP assay, and can lead to false negatives. The use of a second fluoro-label in hit confirmation avoids the loss of false negatives, but still includes false positives.



Screening of the hits against the second peptide site, E7, identified non-specific inhibitors, which caused gross structural changes to the protein. These were excluded from further biochemical testing. Identification of these non-specific inhibitors dramatically reduced the down stream work load.

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The developed screening strategy rapidly identifies false negatives and positives (interfering and protein unfolding reagents) from the primary screen. This reduces the number of compounds to test in biochemical assays, thus saving both time and reagents which will accelerate the hit to lead discovery process.

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ANS (aniline naphthalene sulphonic acid) reagent may be used to detect hydrophobic surfaces on pRb which become exposed as it unfolds. The fluorescence of ANS is highly sensitive to its environment. In solution there is virtually no fluorescence, whereas when bound to protein, such as pRb, it fluoresces highly. Changes in protein structure can alter the fluorescent signal of bound ANS due to changes in its environment to water. Therefore changes in pRb structure can be detected on addition of ANS and the agent that modulates pRb/E2F<sub>(409-426)</sub> complex. If the fluorescent signal alters on addition of the agent, the agent may be affecting the pRb structure. The use of ANS to monitor protein unfolding is known in the art (Semisotnov et al (1991) Biopolymers, 31(1), 119-128)

Biochemical assays could include IC50, isothermal calorimetry, and/or co-crystallisation.

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In an example of an IC50 assay, reactions were performed in 96-well black plates in a total reaction volume of 100  $\mu$ L. Compounds were dissolved in DMSO at a maximum concentration of 10 mM and doubling dilutions made in DMSO. 4  $\mu$ L of diluted compound was mixed with 80  $\mu$ L pRb (400 nM in binding buffer). The plate was incubated at room temperature for 15 min and then Rhodamine-E2F and fluorescein-



E7 were added to give final concentrations of 400 nM each. Reactions were performed in triplicate. Plates were read after 1 hr. The results are shown in Figures 9a and 9b.

- 5 Accordingly, an assay method could include the following steps:
  - a) allow complex formation of pRb and E2F<sub>(409-426)</sub>-fluoropeptide, and measure maximum fluorescence polarization; and
  - b) add a selected agent and detect whether there is a decrease in fluorescence polarization.

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Alternatively, an assay method could include the steps:

- a) allow complex formation of pRb and E2F<sub>(409-426)</sub>-fluoropeptide in the presence and absence of a selected agent and measure the fluorescence polarization; and
- b) compare the fluorescence polarization values.

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Compounds which stabilise the pRb/E2F $_{(409-426)}$  complex could be assessed in a modification of the above method, involving competition binding of pRb by unlabelled E2F $_{(409-426)}$  and E2F $_{(409-426)}$ -fluoropeptide.

- 20 In this regard an assay method could include the following steps:
  - a) allow complex formation of pRb/E2F<sub>(409-426)</sub>-fluoropeptide, and measure max fluorescence polarization;
  - b) add a selected agent and measure fluorescence polarization if no change in fluorescence polarization there is no disruption of complex;
- 25 c) add unlabeled E2F<sub>(409-426)</sub> and measure fluorescence polarization expect displacement of E2F<sub>(409-426)</sub>-fluoropeptide and a decrease in fluorescence polarization, but not if complex is stabilised by presence of the agent.

Alternatively, the pRb, E2F<sub>(409-426)</sub>-fluoropeptide and agent could be added together 30 before detecting fluorescence polarization. If fluorescence polarization is reduced to

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less than a predetermined value, the agent is determined to destabilize the complex, and vice versa.

The interactions could be confirmed by co-crystalisation of pRb/E2F<sub>(409-426)</sub> with the agent, and determination of the three dimensional atomic coordinates by X-ray diffraction and molecular replacement.

The E2F<sub>(409-426</sub>/pRb interaction can also be applied to heterogeneous assay formats (i.e. ones in which reagents are partitioned between a solid support and in solution, and wash steps are involved). This would involve the immobilisation of pRb on microtitre plates, for example by antibody capture or metal ion chelation using Histagged pRb and Nickel coated plates. E2F<sub>(409-426)</sub> peptide may be tagged with fluorescence as above and the fluorescence detected directly to determine binding. Alternatively, the peptide could be labelled with biotin and detected with streptavidinhorse radish peroxidase in an ELISA-like format.

Compounds which interact with the complex without altering association or disassociation of the complex could be identified by crystallographic means, unless the agent itself was tagged (radioactivity/fluorescence) and binding to the complex measured directly, eg fluorescence polarization as above or scintallation counting of an immuno-precipitate.

Alternatively, the agent can be added to pRb and  $E2F_{(409-26)}$  under conditions in which pRb and  $E2F_{(409-26)}$  can form a complex. This could result in the agent and complex cocrystallising. The binding affinities of pRb to  $E2F_{(409-26)}$  in the pRb/  $E2F_{(409-26)}$  complex in the presence and absence of the agent can then be compared to determine whether the agent inhibits complex formation. The three dimensional structure of the pRb/  $E2F_{(409-26)}$  – agent complex can also be solved (X-ray diffraction using molecular replacement analysis) to enable the associations in the new complex to be compared with those in the pRb/  $E2F_{(409-26)}$  complex (see Annex 1). As a further alternative the



pRb/  $E2F_{(409-26)}$  complex can be formed before soaking the complex in the presence of a selected agent. The binding affinities of pRb to  $E2F_{(409-26)}$  can then be determined in the presence and absence of the agent. As before, the three dimensional structure of any pRb/  $E2F_{(409-26)}$  – agent complex could be solved.

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The binding affinities can be measured using isothermal titration calorimetry.

Alternatively, surface plasmon resonance (SPR) could be used such as that provided by Biacore AB.

Preferred features of each aspect of the invention are as for each of the other aspects mutatis mutandis. The prior art documents mentioned herein are incorporated to the fullest extent permitted by law.

## Annex 1

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719		LEU	A 468	3.06	.97	5.57	.00 23.4
720		LEU	A 468	3.8	7.109	5.98	.00 23.
721		LEU	A 468	4.09	.05	.80	.00 23.4
722		LEU		4.92	.68	œ	.00 23.3
723		LEU	A 468	4.39	.45	.39	.00 19.9
724		LEU	A 468	6.38	.04	ω.	.00 25.5
725		LEU	A 468	3.17	.91	0.	.00 24.2
726		LEU		m.	w	47.608	23.4
727		SER		1.94	.54	•	.00 24.4
728		SER	A 469	1.16	.35	.31	7
729				1.89	8.580	49.621	26.2
730				2.03	.35	.33	.00 24.9
731		-	A 469	۰.	9.644	47.500	1.00 26.91
132		-		1.2	62	.28	.00 28.4
733		-		99.0	0.77	9.	.00 26.9
34				ö	11.941	_	.00 25.8
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38				•	.31	.94	.00 25.9
33		•		•	1.81	.26	25.8
40		•		•	.76	45.661	23.6
41		•	A 471		12.878	.17	26.3
42					.89	.21	.00 27.7
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44				15.514	12.376	46.723	1.00 32.75
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46		•	A 471		•	48.845	36.4
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49			A 471	.17		-	0 28.3
20		ASN 1	A 472	7.16	4.	ů.	0 25.5
21			A 472	7.75	5.73	.68	.00 25.1
152		ASN 1	A 472	ω.	6.42	43.345	4.9
753		ASN 1	A 472	9.34	7.80	.70	.00 28.3
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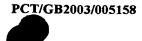
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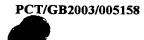


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**SUBSTITUTE SHEET (RULE 26)** 

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SUBSTITUTE SHEET (RULE 26)

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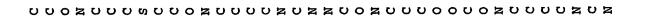
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50.129		•	•	48.418	49.824	50.774	50.583	51.722	46.173	45.826	45.346	43.892	43.246	43.781	43.456	43.952	44.790	45.273	45.130	44.643	43.456	42.342	44.334	43.993	45.024	44.958	43.629	43.221	42.974	43.839	43.184	44.370	44.294	45.194	46.133	46.625
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**SUBSTITUTE SHEET (RULE 26)** 



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1.293	2.492	•	2.688	2.596	1.332	•	0.077	3.627	•	4.874	5.925	7.200	8.469	9.744	10.900	11.587	11.314	12.606	5.569	•	5.417	5.019	6.024	7.416	•	•	•	3.257	•	•	•	.67	.30	2.334	.26	•
47.261	2.53	42.079	40.705	40.166	40.634	39.938	œ	39.780	38.662	40.219	39.358	40.136	39.302	40.166	39.304	39.133	39.826	38.279	38.683	39.321	37.374	36.620	36.851	36.238	35.077	36.823	36.989	36.893	37.458	37.862	39.393	٥,	4	•	42.863	42.877
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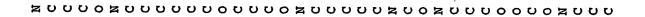
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**SUBSTITUTE SHEET (RULE 26)** 

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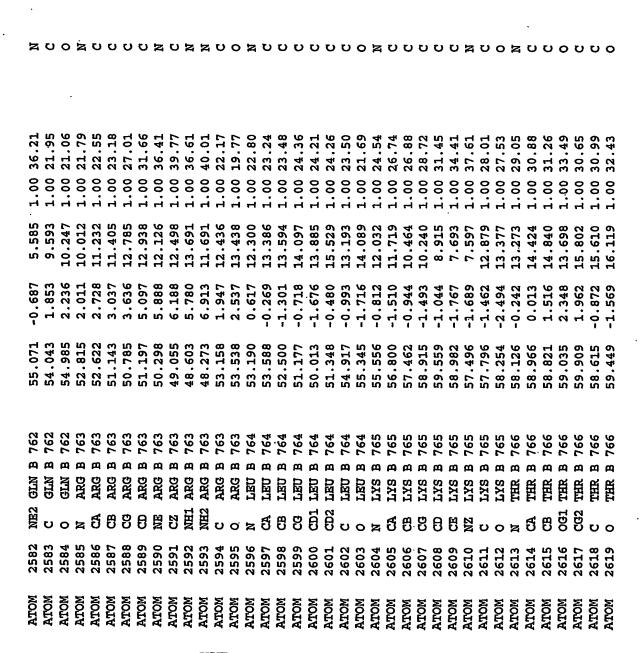
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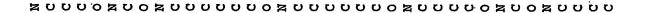
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37.125	6.59	6.30	5.76	6.53	7.06	9.96	.91	0.49	1.1	.56	.94	2.68	3.49	90.	0.60	53	1.59	42.277	.68	2.34	.13	. 14	•	•	42.606	•	•	•	.98	.80	.21	4.	.01	5.97		6.83	7.87
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SUBSTITUTE SHEET (RULE 26)





ATOM ATOM	2620 2621	ූ ව්	ASN	д д	767 767	57.398	-0.804	16.083	1.00 3	1.61
	2622	8	ASN	M	767	9.	02	7.7	.00	9
ATOM	62	ខ្ល	ASN	Д	767	5.04	æ	8.83	.00 3	
ATOM	2624	9	ASN	ф	167	₹1	à	8.54	.00 3	ų.
ATOM	2625	ND2	ASN	ф	167	5.14	.45	0.09	.00 4	9
	2626	ပ	ASN	ф	167	5.92	.09	16.985	.00	ល
	2627	0	ASN	М	767	7.39	æ	.81	.00	7
ATOM	2628	×	ILE ILE	ф	768	56.354	-3.511	15.863	1.00 3	5.47
	2629	ජ	田田	ф	768	23	-4.940	15.625	1.00 3	6.32
	2630	8	ILE	ф	168	5.62	Ġ	4.26	0 3	4.
ATOM	w	CG1	ITE	ф	768	4.10	σ.	.30	.00 3	7
	53	មួ	ILE	щ	768	3.42	-5.168	•	.00 3	4.
ATOM	w	CG2	II.	ф	768	. 95	•	13.834	.00	4.18
ATOM	53	บ	ILE	ф	768	57.592	-5.577	•	.00 3	7.63
	53	0	HE	ф	768	7.69	-6.786	.92	1.00 3	8.19
ATOM	2636	z	LEU	m	169	. 62	-4.764	15.505	1.00 3	8.55
	เก	ජ	LEU	æ	769	59.996	-5.257	15.523	1.00 4	0.19
	2638	ප	LEG	m	769	60.996	-4.186	15.082	1.00 4	66.0
ATOM	2639	පු	LEU	ф	769	61.863	-4.615	13.896	1.00 4	2.49
	54	9	LEU	ф	769	.46	-5.984	.25	1.00 4	5.30
ATOM	2641	9	LEU	Ø	169	61.061	-4.723	•	1.00 42	ø.
ATOM	64	ບ	LEU	æ	769	0.37	.75	•	0 4	o,
ATOM	64	0	LEU	ф	169	1.02	-6.789	9	0 4	
ATOM	49	z	GLN	æ	770	.98	-5.024	17.936	1.00 4	1.51
ATOM	2645	ජ	GLN	ф	770	.28	-5.473	19.304	.00 4	1.74
ATOM	2646	ප	GLN	ф	770	59.366	-4.794	ü	1.00 4	1.53
ATOM	2647	ន	GLN	ф	770		-5.418	21.762	_	•
ATOM	2648	₿	GIN	ф	770	8.13	•	ĸ	1.00 42	2.74
ATOM	2649	<b>OE1</b>	GLN	ф	770	90.	-6.041	23.668	1.00 4	1.16
ATOM	2650	NE2	GLN	ф	770	57.116	-4.740	0	1.00 4	3.74
ATOM	2651	ບ	GLN	œ	770	60.125	-7.004	19.380	1.00 4	1.82
ATOM	65	0	GLN	ø	770	61.077	-7.747	•	0 4	1.81
ATOM		×	TXR	'n	771	.91	4.	4	1.00 4	2.24
ATOM	9	ජ	TYR		771	9	œ	.22	.00	ö
ATOM	2655	巴	TXR.	д	771	•	4	ω.	4	œ
ATOM	2656	පු	TX	ø	771	6.1	H	.35	.00 3	•
ATOM	65	9	TXR	B	771	5.7	0	0.66	.00 3	m.



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PRO B 2707 2708 2708 2710 2711 2712 2713 2714 2714 2717 2719 2719 2720 2721 2702 2703 2704 2705 2706 2724 2725 2726 2727 

41.11 51.46 53.83 54.70 54.84 53.76 42.35 50.08 51.26 51.67 48.44 49.03 111111 00001 00001 1.00 29.879 31.708 32.656 33.902 35.137 35.248 37.122 31.879 31.746 31.075 29.563 28.831 31.643 29.579 27.311 29.341 28.388 29.710 30.932 28.374 30.413 28.967 32.048 -9.692 -10.450 -11.490 -10.203 -9.990 -9.080 -12.473 -12.667 -10.093 -8.777 -9.749 -9.334 -11.196 -10.507 -11.356 -12.545 -13.068 -12.254 -11.180 -12.109 -13.299 -12.296 -12.469 -13.007 39.388 38.831 38.870 38.178 39.185 39.506 39.720 38.331 37.169 36.409 35.162 35.332 37.583 44.169 45.321 42.915 40.617 40.666 41.830 40.049 35.757 42.817 45.027 2748 2749 2751 2751 2751 2753 2753 2755 2755 2756 2756 2760 2760 2760 2760 2760 2739 2742 2744 2745 2746 2747 2764 2765 2767 ATOM ATOM ATOM ATOM

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10.542	11.351	12.403	13.079	12.312	12.999	12.033	12.741	12.001	14.012	13.540
24.662	25.144	27.470	26.583	28.717	29.152	29.110	29.079	29.132	29.007	30.566
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1. A crystal structure of the pRb/E2F<sub>(409-426)</sub> complex, characterised by the atomic coordinates of Annex 1.

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2. A crystal structure as claimed in claim 1, wherein the interactions between E2F<sub>(409)</sub>.

426) and pRb comprise one or more of the following interactions:

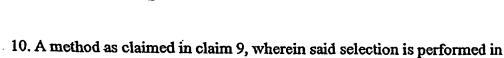
E2F <sub>(409-426)</sub> residue	pRb residue
Leu <sub>409</sub>	Lys <sub>548</sub>
Tyr <sub>411</sub>	Glu <sub>551</sub>
Tyr411	Ile <sub>532</sub>
Тут411	Glu <sub>554</sub>
His <sub>412</sub>	Arg <sub>656</sub>
His <sub>412</sub>	Lys <sub>653</sub>
Gly <sub>414</sub>	Glu <sub>533</sub>
Gly <sub>414</sub>	Lys <sub>652</sub>
Leu <sub>415</sub>	Leu <sub>649</sub>
Leu <sub>415</sub>	Glu <sub>553</sub>
Leu <sub>415</sub>	Lys <sub>537</sub>
Glu <sub>417</sub>	Lys <sub>537</sub>
Gly <sub>418</sub>	Arg <sub>467</sub>
Glu <sub>419</sub>	Thr <sub>645</sub>
Arg <sub>422</sub>	Glu <sub>464</sub>
Asp <sub>423</sub>	Arg <sub>467</sub>
Leu <sub>424</sub>	Lys <sub>530</sub>
Phe <sub>425</sub>	Phe <sub>482</sub>
Phe <sub>425</sub>	Lys475
	. <del>  </del>

- 3. A method to identify an agent which modulates the interaction between pRb and E2F<sub>(409,426)</sub>, the method comprising:
- a) combining together pRb, E2F<sub>(409-426)</sub> and an agent, under conditions in which pRb
   and E2F<sub>(409-426)</sub> form a complex;
  - b) growing a crystal structure of any pRb/ E2F<sub>(409-426)</sub> complex; and
- c) analysing the crystal to determine whether the agent is an agent which modulates
  the interaction between pRb and E2F<sub>(409-426)</sub>.
  - 4. A method, as claimed in claim 3, wherein the combining of the components is pRb with the agent and then  $E2F_{(409-426)}$ .
- 5. A method as claimed in claim 3, wherein the combining of the components is  $E2F_{(409-426)}$  with the agent and then pRb.
  - 6. A method as claimed in claim 3, wherein the combining of the components is pRb with E2F<sub>(409-426)</sub> and then the agent.

- 7. A method as claimed in any one of claims 3 to 6, wherein step c) comprises comparing the crystal structure to the crystal structure of claim 1
- 8. A method as claimed in any one of claims 3 to 7, wherein the agent is selected using the three dimensional atomic co-ordinates of Annex 1.
  - 9. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising selecting an agent using the three-dimensional atomic coordinates of Annex 1.

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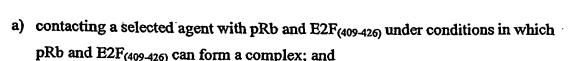
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- conjunction with computer modeling.
- 11. A method as claimed in claim 9 or 10, wherein the method further comprises the steps of:
  - a) contacting the selected agent with pRb and E2F<sub>(409-426)</sub> under conditions in which pRb and E2F<sub>(409-426)</sub> can form a complex; and
- b) measuring the binding affinity of pRb to E2F<sub>(409-426)</sub> in the presence of the agent and comparing the binding affinity to that of pRb to E2F<sub>(409-426)</sub> when in the
   absence of the agent, wherein an agent modulates a pRb/E2F<sub>(409-426)</sub> complex when there is a change in the binding affinity of pRb to E2F<sub>(409-426)</sub> when in the presence of the agent.
  - 12. A method as claimed in claim 11, wherein the method further comprising:
- a) growing a supplementary crystal from a solution containing pRb and E2F<sub>(409-426)</sub> and the selected agent where said agent changes the binding affinity of the pRb/E2F<sub>(409-426)</sub> complex under conditions in which pRb and E2F<sub>(409-426)</sub> can form a complex;
  - b) determining the three-dimensional atomic coordinates of the supplementary crystal by X-ray diffraction using molecular replacement analysis;
  - c) comparing the three dimensional coordinates with those for the crystal structure as claimed in claim 1; and
  - d) selecting a second generation agent using the three-dimensional atomic coordinates determined for the supplementary crystal.
  - 13. A method as claimed in claim 12, wherein said selection is performed in conjunction with computer modeling.
  - 14. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:

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- b) measuring the binding affinity of pRb to E2F<sub>(409-426)</sub> in the presence of the agent and comparing the binding affinity to that of pRb to E2F<sub>(409-426)</sub> when in the absence of the agent, wherein an agent modulates a pRb/E2F<sub>(409-426)</sub> complex when there is a change in the binding affinity of pRb to E2F<sub>(409-426)</sub> when in the presence of the agent.
- 15. A method as claimed in claim 14, wherein the method further comprising:
- a) growing a supplementary crystal from a solution containing pRb and E2F<sub>(409-426)</sub> and the selected agent where said agent changes the binding affinity of the pRb/E2F<sub>(409-426)</sub> complex under conditions in which pRb and E2F<sub>(409-426)</sub> can form a complex;
  - b) determining the three-dimensional atomic coordinates of the supplementary crystal by X-ray diffraction using molecular replacement analysis;
  - c) comparing the three dimensional coordinates with those for the crystal structure claimed in claim 1; and
  - d) selecting a second generation agent using the three-dimensional atomic coordinates determined for the supplementary crystal.
  - 16. A method as claimed in claim 15, wherein said selection is performed in conjunction with computer modeling.
  - 17. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:
    - a) selecting an agent;
    - b) co-crystalising pRb with the agent;
    - c) determining the three dimensional coordinates of the pRb-agent association by X-ray diffraction using molecular replacement analysis; and



- d) comparing the three dimensional coordinates with those of the crystal structure claimed in claim 1.
- 18. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:
- a) selecting an agent;
- b) crystalising pRb and soaking the agent into the crystal;
- c) determining the three dimensional coordinates of the pRb-agent association by X-ray diffraction using molecular replacement analysis; and
- d) comparing the three dimensional coordinates with those of the crystal structure claimed in claim 1.
  - 19. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:
- 15 a) selecting an agent;
  - b) co-crystalising pRb, E2F<sub>(409-426)</sub> and the agent;
  - c) determining the three dimensional coordinates of the pRb-E2F-agent association by X-ray diffraction using molecular replacement analysis; and
  - d) comparing the three dimensional coordinates with those of the crystal structure claimed in claim 1.
  - 20. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:
  - a) selecting an agent;
- b) co-crystalising pRb and E2F<sub>(409-426)</sub> and soaking the agent into the crystal;
  - c) determining the three dimensional coordinates of the pRb-E2F-agent association by X-ray diffraction using molecular replacement analysis; and
  - d) comparing the three dimensional coordinates with those of the crystal structure claimed in claim 1.



- 21. A method as claimed in any one of claims 17 to 20, wherein the agent is selected using the three dimensional atomic co-ordinates of Annex 1
- 22. A method as claimed in any one of claims 17 to 21, wherein the methods further
  comprise selecting a second generation agent using the three dimensional atomic coordinates determined in step c).
  - 23. A method as claimed in claim 22, wherein the second generation agent is selected using the three dimensional atomic coordinates of Annex 1.
  - 24. A method as claimed in claim 22 or 23, wherein the selection is performed in conjunction with computer modeling.
- 25. A method of identifying an agent as claimed in any one of claims 3 to 24, wherein the selected agent and/or the second generation agent mimics a structural feature of E2F<sub>(409-426)</sub>, when said E2F<sub>(409-426)</sub> is bound to pRb.
  - 26. A method as claimed in claim 9 or 10, wherein method comprises the further steps of:
- a) contacting the selected agent with a pRb/E2F<sub>(409-426)</sub> complex; and
  - b) determining whether the agent affects the stability of the complex.
  - 27. A method as claimed in claim 26, wherein the determination is with fluorescence polarization.

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- 28. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising:
- a) contacting a fluorescently tagged E2F<sub>(409-426)</sub> peptide (E2F-fluoropeptide) with pRb to allow pRb/E2F-fluoropeptide complex formation;
- 30 b) detecting the fluorescence polarization;



- c) adding a selected agent; and
- d) detecting the fluorescence polarization in the presence of the agent.
- 29. A method of identifying an agent that modulates a pRb/E2F<sub>(409-426)</sub> complex, comprising;
- a) contacting a fluorescently tagged E2F<sub>(409-426)</sub> peptide (E2F-fluoropeptide) with pRb to allow pRb/E2F-fluoropeptide complex formation;
- b) detecting the fluorescence polarization;
- c) contacting a selected agent with pRb and E2F<sub>(409-426)</sub> peptide (E2F-fluoropeptide) under conditions in which pRb and E2F-fluoropeptide can form a complex;
  - d) detecting the fluorescence polarization; and
  - e) comparing the fluorescence polarization detected in b) and d).
- 30. A method as claimed in claim 28 or 29, wherein the fluorescently tagged E2F peptide is selected using the three dimensional atomic co-ordinates of Annex 1.
  - 31. A method as claimed in any one of claims 28 to 30, wherein a decrease in fluorescence polarization in the presence of the agent indicates that the agent destabilises the complex.

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- 32. A method as claimed in any one of claims 28 to 31, wherein the method comprises the further step of adding untagged E2F<sub>(409-426)</sub> and detecting fluorescence polarization.
- 25 33. A method as claimed in claim 32, wherein if fluorescence polarization decreases, on addition of the untagged E2F<sub>(409-426)</sub>, the agent does not stabilise the complex.
  - 34. A method as claimed in claim 32 or 33, wherein if there is no substantial change in fluorescence polarization, on addition of the untagged E2F<sub>(409-426)</sub>, the agent stabilises the complex.

- 35. A method as claimed in any one of claims 28 to 34, wherein the method further comprises:
- a) contacting a fluorescently tagged E7 peptide (E7-fluoropeptide) with pRb to allow
   pRb/E7-fluoropeptide complex formation;
  - b) detecting the fluorescence polarization;
  - c) adding an agent that modulates pRb/E2F(409-426) complex; and
  - d) detecting the fluorescence polarization in the presence of the agent.
- 36. A method as claimed in any one of claims 28 to 34, wherein the method further comprises:
  - a) contacting a fluorescently tagged E7 peptide (E7-fluoropeptide) with pRb to allow pRb/E7-fluoropeptide complex formation;
  - b) detecting the fluorescence polarization;
- c) contacting an agent that modulates pRb/E2F<sub>(409-426)</sub> complex with pRb and E7-fluoropeptide under conditions in which pRb and E7-fluoropeptide can from a complex;
  - d) detecting the fluorescence polarization; and
  - e) comparing the fluorescence polarization detected in b) and d).

- 37. A method as claimed in claim 35 or 36, wherein a decrease in fluorescence polarization indicates that the agent also inhibits E7 binding to pRb.
- 38. A method as claimed in any one of claims 11 to 16, wherein the binding affinity is measured by isothermal titration calorimetry.
  - 39. A method as claimed in any one of claims 11 to 16, wherein the binding affinity is measure by Surface Plasmon Resonance (SPR).

- 40. An agent, that modulates the interaction between pRb and E2F<sub>(409-426)</sub>, identified by a method as claimed in any one of claims 3 to 39.
- 41. An agent, as claimed in claim 40, for use as an apoptosis promoting factor in the prevention or treatment of proliferative diseases.
  - 42. An agent as claimed in claim 40 or 41, wherein the agent is for use in preventing or treating cancer, which may be pancreatic cancer and related diseases.
- 43. The use of an agent, which modulates the formation of a pRb/E2F<sub>(409-426)</sub> complex, identified by a method as claimed in any one of claims 3 to 39, in the manufacture of a medicament for the prevention or treatment of proliferative diseases.
- 44. The use of an agent as claimed in claim 43, wherein the proliferative diseases are cancer, preferably pancreatic cancer and related diseases.
  - 45. The use of the atomic co-ordinates of the crystal structure as claimed in claim 1 or 2, for identifying an agent that modulates the formation of a pRb/E2F<sub>(409-426)</sub> complex.
  - 46. Computer readable media comprising a data storage material encoded with computer readable data, wherein said computer readable data comprises a set of atomic co-ordinates of the pRb/E2F<sub>(409-426)</sub> crystal structure of Annex 1 recorded thereon.

FIG. 1A

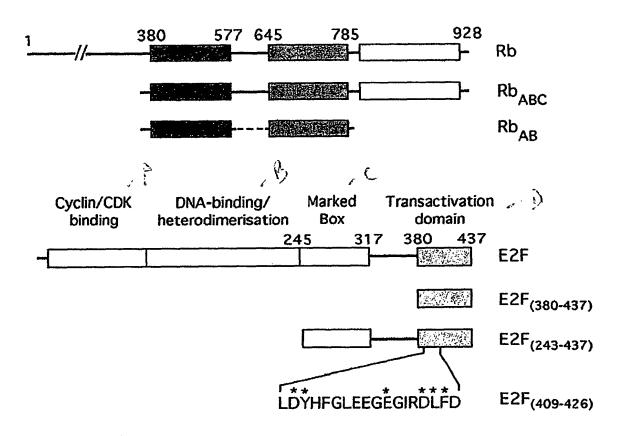
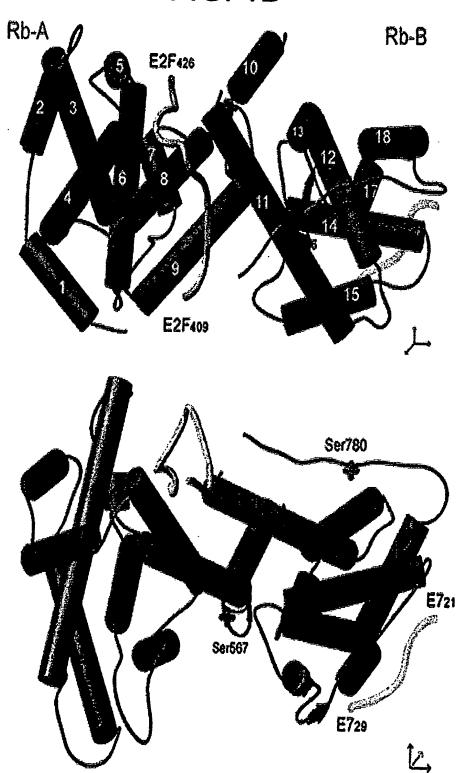


FIG. 1B



**SUBSTITUTE SHEET (RULE 26)** 





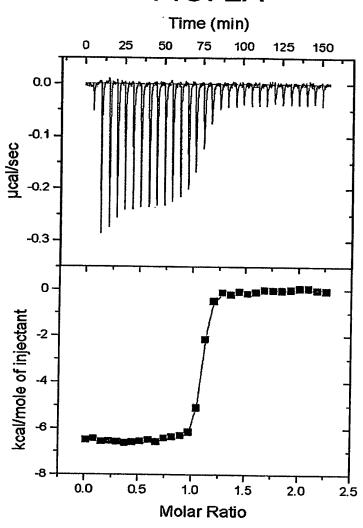


FIG. 2B

Binding Constants (μΜ)	Rb <sub>AB</sub>	Rb <sub>ABC</sub>
E2F (409-426)	0.34 <u>+</u> 0.02	0.3 <u>+</u> 0.03
E2F (380-437)	0.16 <u>+</u> 0.01	0.1 <u>+</u> 0.01
E2F (243-437)	<0.01	<0.01

FIG. 3
Binding of Fluorescein-E2F, Rhodamine-E2F and Fluorescein-E7 to pRb

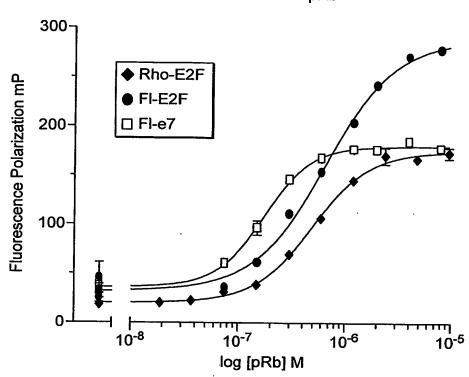
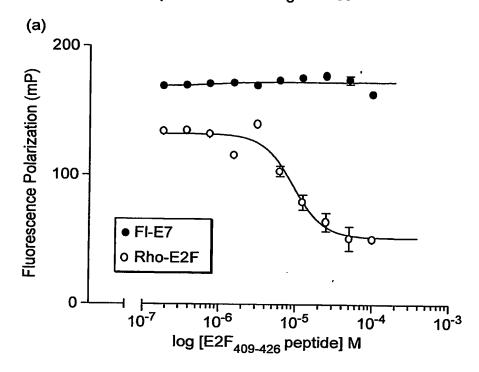


FIG. 4
Displacement Binding Curves



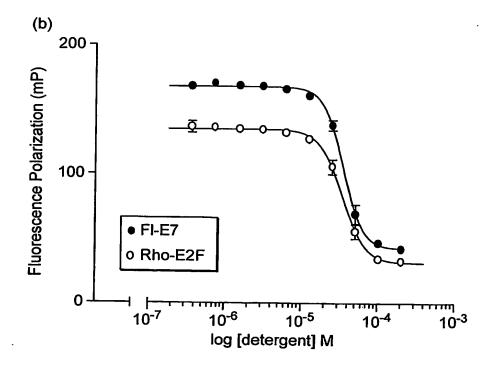


FIG. 5
Screen controls from Test Screen 6X384 plates

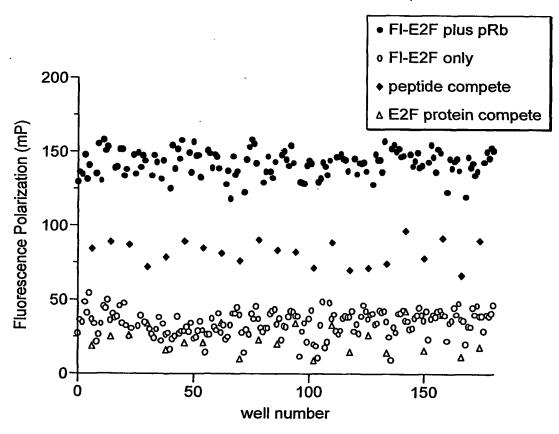


FIG. 6
Correlation Inhibition Rhodamine and Fluorescein-E2F

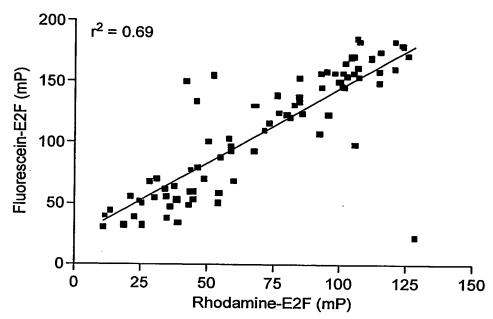


FIG. 7
Correlation Inhibition Fluorescein-E2F and Fluorescein-E7

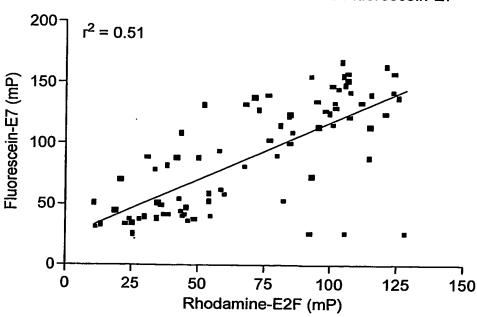
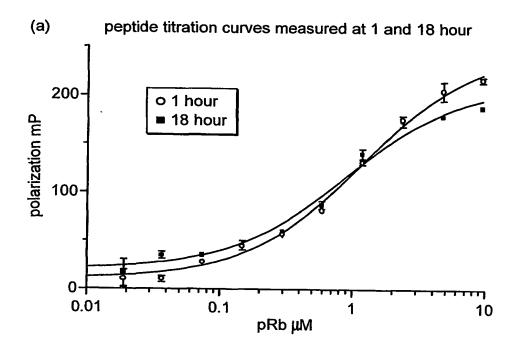
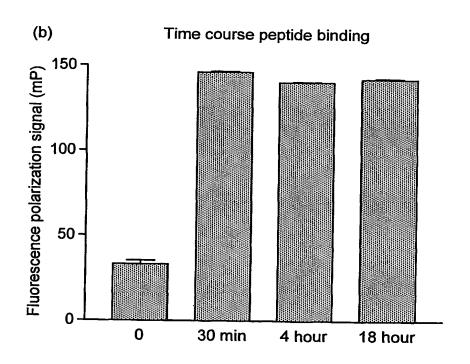


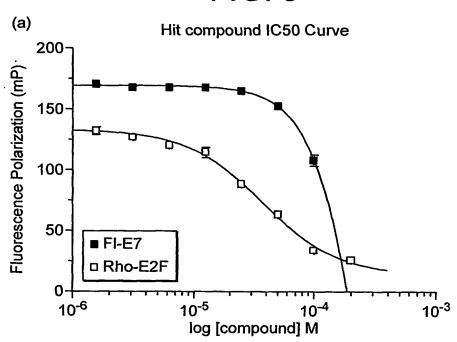
FIG. 8

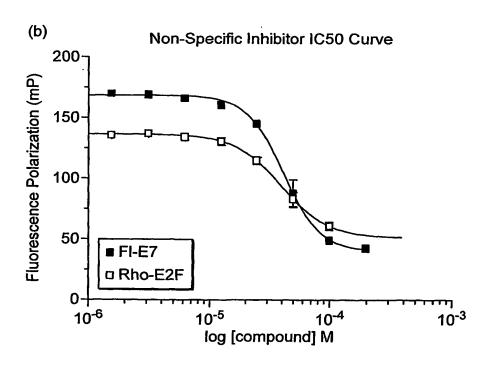




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FIG. 9





# INTERNATION SEARCH REPORT

Internati Application No PCT/B/05158

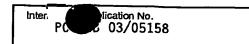
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A CLASSI IPC 7	CO7K14/47			
According to	o International Patent Classification (IPC) or to both national classif	ication and IPC		
	SEARCHED			
Minimum do IPC 7	ocumentation searched (classification system followed by classification CO7K	ation symbols)		
Documenta	tion searched other than minimum documentation to the extent that	such documents are inclu	ided in the fields so	sarched
	iata base consulted during the International search (name of data b , EPO-Internal, MEDLINE	ase and, where practical,	search terms used	0
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT			
Category °	Citation of document, with indication, where appropriate, of the n	elevant passages		Relevant to claim No.
Υ	LEE JIE-OH ET AL: "Structure of retinoblastoma tumour-suppressor domain bound to a peptide from H NATURE (LONDON), vol. 391, no. 6670, 26 February 1998 (1998-02-26), p 859-865, XP002272473 ISSN: 0028-0836 the whole document	pocket PV E7"		3-39,45
Y	HELIN K ET AL: "A CDNA ENCODING PRB-BINDING PROTEIN WITH PROPERT TRANSCRIPTION FACTOR E2F" CELL, CELL PRESS, CAMBRIDGE, NA, vol. 70, no. 2, 24 July 1992 (19 pages 337-350, XP000872846 ISSN: 0092-8674 the whole document	IES OF THE		3-39,45
χ Furth	ner documents are listed in the continuation of box C.	Patent family m	embers are listed in	n annex.
"A' docume conside a filling de filling fillin	nt which may throw doubts on priority claim(s) or solited to establish the publication date of another or or other special reason (as specified) and referring to an oral disclosure, use, exhibition or neans and published prior to the international filling date but an the priority date claimed actual completion of the international search	"Y" document of particul cannot be considered cocument is combinents, such combinents, such combinents, such combinents, such combinents art.  "&" document member of the particular of the part	ithe principle or the lar relevance; the cleed novel or cannot e step when the doc ar relevance; the cleed to involve an inv ned with one or mo- nation being obviou of the same patent for e international sear	the application but tory underlying the laimed invention be considered to sument is taken alone laimed invention entive step when the re other such docusis to a person skilled
	A April 2004  Halling address of the ISA  European Patent Office, P.B. 5818 Patentlaan 2  NL - 2280 HV Rijswijk  Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  Fax: (+31-70) 340-3016	22/04/20 Authorized officer Wimmer,		

# INTERNAMINAL SEARCH REPORT

Internat pplication No PCT/ 05158

		PCT//05158
	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	DATABASE PDB 'Online! RCSB; 7 January 2003 (2003-01-07) LEE CHANGWOOK ET AL: "Structure of rb tumor suppressor bound to the transactivation domain of e2f-2" retrieved from PDB Database accession no. 1n4m XP002272474 the whole document & LEE CHANGWOOK ET AL: "Structural basis for the recognition of the E2F transactivation domain by the retinoblastoma tumor suppressor." GENES & DEVELOPMENT. UNITED STATES 15 DEC 2002, vol. 16, no. 24, 15 December 2002 (2002-12-15), pages 3199-3212, ISSN: 0890-9369	3-39,45
P,X	DATABASE PDB 'Online! RCSB; 6 March 2003 (2003-03-06) XIAO ET AL: "Crystal Structure Of The Retinoblastoma Tumour Suppressor Protein Bound To E2F Peptide" retrieved from PDB Database accession no. 109k XP002272475 abstract & XIAO ET AL.: "Crystal Structure of the Retinoblastoma Tumor" PROC.NATL.ACAD.SCI., vol. 100, no. 5, 21 February 2003 (2003-02-21), pages 2363-2368, USA	3-39,45





Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This inte	emational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: 1 2 46 because they relate to subject matter not required to be searched by this Authority, namely:
	Rule 39.1(v) PCT - Presentation of information
2. X	Claims Nos.: 40-44 because they relate to parts of the International Application that do not comply with the prescribed requirements to such a carried out, specifically:
	see FURTHER INFORMATION sheet PCT/ISA/210
з. 🗌	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This inter	mational Searching Authority found multiple inventions in this international application, as follows:
!	
1.	As all required additional search fees were timely paid by the applicant, this international Search Report covers all searchable claims.
2	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. 🔲 !	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark o	The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.

### FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 40-44

Present claims 40-44 relate to compounds defined by reference to a desirable characteristic or property, namely that they modulate the interaction between pRb and E2F, and that they may be identified by one of the claimed in silico sceening or modeling methods.

The claims cover all compounds having this characteristic or property, whereas the application provides support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT for no such compounds. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claims also lack clarity (Article 6 PCT). An attempt is made to define the compounds by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, no search has been carried out for these claims.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

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